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Page 002

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Baron Zieles

Name

April 11, 2005

Date

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Inventors: Thomas R. Cech et al.

Art Unit: 1632

Filing Date: January 11, 2002

Examiner: Louis D. Lieto, Ph.D.

Serial No: 10/044,539

Docket: 015389-002630US; 018/212c

Title: MAMMALIAN CELLS THAT HAVE
INCREASED PROLIFERATIVE CAPACITY

DECLARATION UNDER 37 CFR § 1.132

BY GREGG B. MORIN, Ph.D.

Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

I, GREGG B. MORIN, do hereby declare as follows:

1. I am a co-inventor on the above-referenced patent application, and am familiar with its contents. I have been a research scientist in the field of molecular biology for about twenty years, and am currently Head of Proteomics at the Michael Smith Genome Sciences Centre in Vancouver, British Columbia.

PATENT
10/044,539
Docket 018/212

2. The claims currently under examination at the Patent Office cover mammalian cells containing a recombinant polynucleotide encoding human telomerase reverse transcriptase (hTRT). I understand the Patent Office has raised the question of whether someone reading the disclosure at the time it was filed would have known how to make fragments of the hTRT protein encoded by SEQ. ID NO:1 that would still have telomerase catalytic activity when associated with telomerase RNA component.

3. It is standard procedure in determining the structure-function relationship of any protein to truncate the sequence from either terminus until there is loss in function. The minimal fragment that retains function defines the core sequence needed for activity.

4. I enclose two scientific papers describing the use of terminal deletion analysis to map functional domains within a protein sequence. The first paper by Cherry et al. (Plant Cell 5:563-575, 1993) maps the functional regions of oat Phytochrome A. The second paper by Kawakami et al. (Nucl. Acids Res. 24:303-310, 1996) maps the functional regions of mouse homeobox protein ARRC3. These papers were published some time before the filing of our first hTRT patent application, showing that fragmentation analysis was a well-known and often used strategy of protein analysis.

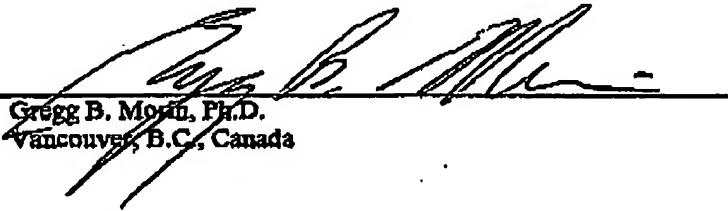
5. In a similar fashion, the core sequence of hTRT could be determined by fragmentation analysis in combination with functional testing. Once the core sequence is identified, fragments of hTRT smaller than the entire full-length protein but containing the core sequence would be expected to retain function. Smaller versions of the native protein that retain activity may be more easy to crystallize. They may be better for use in vitro, in vivo, or for biomedical applications because other activities or functions normally mediated by the deleted part of the sequence have been eliminated.

PATENT
10/044,539
Docket 018/212

6. One way active hTRT fragments could be found would be to start with an expression plasmid like pGRN121, described in Example 6 (SEQ. ID NO:1 in the EcoRI site of pBluescriptISK+). One or more codons can be removed by recombinant techniques from either end of the hTRT encoding sequence, thus creating a vector for expressing the truncated protein. The vector could then be expressed in a reticulocyte lysate system along with hTR (telomerase RNA component) and assayed for telomerase activity using the same methodology shown in Example 7.
7. In my expert opinion, someone skilled in recombinant DNA technology as it was in 1997 and guided by the information in this patent application would have been able to perform this type of fragmentation study as a matter of routine experimentation.
8. I hereby declare that all statements made in this Declaration of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

7 April 2004

Date



Gregg B. Modlin, Ph.D.
Vancouver, B.C., Canada